

Isolation, Characterization and Identification of Bacterial isolates from Auto-mechanic Workshop contaminated with Hydrocarbon

Jude O. Osarumwense, Ph.D.¹, Francis A. Igiebor, M.Sc^{2,3*} and Daniel E. Idahosa, B.Sc.¹

¹Department of Science Laboratory Technology, Faculty of Life Sciences, University of Benin, Benin City, Nigeria.

²Department of Microbiology, College of Natural and Applied Sciences, Wellspring University, Benin City, Nigeria.

³Environmental Biotechnology Sustainability Research Group, Department of Plant Biology and Biotechnology, Faculty of Life Sciences, University of Benin, Benin City, Nigeria.

E-mail: francis.igiebor@lifesci.uniben.edu*

ABSTRACT

This work focused on screening of hydrocarbon and heavy metal degrading microorganisms from the soil contaminated with hydrocarbon in auto-mechanic workshop using selective enrichment techniques. It resulted in the collection of 9 distinct species. All strains were cultivated in liquid media with engine oil as a sole carbon and energy source. Bacterial strains capable of degrading hydrocarbons belong to the genera *Bacillus subtilis*, *Escherichia coli*, *Staphylococcus epidermidis*, *Micrococcus latus*, *Staphylococcus aureus*, *Proteus mirabilis*, *Clostridium* spp., *Streptococcus* spp and *Pseudomonas aeruginosa*.

Microorganisms were identified using cultural and morphological characteristics as well as biochemical tests. Rate of biodegradation depends greatly on the composition, state, and concentration of the hydrocarbons. Temperature and oxygen and nutrient concentrations were important variables in the environment. The total hydrocarbon (THC) content on day 2 stood at 139.30 and the amount degraded by *P. aeruginosa* was 0.089 mg/kg. The percentage degraded by the same organism was 8.89 %, compared to THC on day 2 for *B. subtilis* which was 145.70. The amount degraded was 0.047 mg/kg and percentage was 4.71 %. However, *B. subtilis* and *P. aeruginosa* both showed varying level of degradation capabilities.

(Keywords: engine oil, hydrocarbon, microorganisms, degradation, bioremediation, bacteria)

INTRODUCTION

Engine oil is a complex mixture of hydrocarbons and other organic compounds, including some organ-metallic constituents (Butler and Mason, 1997). It is used to lubricate the parts of automobiles engine, in order to keep everything running smoothly (Hagwell *et al.*, 1992). New motor oil contains a higher percentage of fresh and lighter (often more volatile and water soluble) hydrocarbons that would be more of a concern for acute toxicity to organisms. Used motor oil contains more metals and heavy polycyclic aromatic hydrocarbons (PAHs) that would contribute to chronic hazards including mutagenicity and carcinogenicity (Boonchanet *al.*, 2000).

PAHs have a widespread occurrence in various ecosystems that contribute to the persistence of these compounds in the environment (Van Hamme *et al.*, 2003). The illegal dumping of used motor oil is an environmental hazard with global ramifications (Blodgett, 2001). The release of oil into the environment causes environmental concern and attracts the public attention (Rolinget *al.*, 2002). Engine Oil at minimal concentration in soil stimulates growth (Anoliefo and Edegbai, 2000). When microorganisms in the soil come in contact with oil, the initial reaction is a reduction of the activities as a result of reduced air availability (Odu, 1981).

Soil polluted with waste engine oil becomes water logged; inducing several stresses on the plant and microbial community; ranging from changes in structure and configuration of enzymes. Polluted soil could also become

unsuitable due to increase in the toxic levels of elements (Udo and Fayemi, 1975).

The most widely used procedures to degrade waste engine oil in soil are the chemical and physical methods. These methods are however not favourable as they introduce harmful materials into the environment (Davis and Wilson, 2005). The most suitable technology for cleaning spills is the bioremediation method, which must be specific for a particular site; haven met some conditions like the type, quantity and toxicity of contaminant chemicals present and the indigenous microbial population (Ikhajiagbe and Anoliefo, 2011; Osarumwense and Igiebor, 2018).

Other remediation technologies include the addition of nutrient to stimulate the activities of host microbial community. In the presence of favourable environmental condition, there is an increase in the growth of microbial population which results in faster degradation of poisonous materials (Igieboret *al.*, 2017). Some other technologies (phytoremediation and fungal remediation) have been used to clean up polluted soils and underground water (Ikhajiagbe and Anoliefo, 2011).

Bioremediation makes use of indigenous oil-consuming microorganisms, called petrophiles, by enhancing and fertilizing them in their natural habitats. Petrophiles are very unique organisms that can naturally degrade large hydrocarbons and utilize them as a food source (Harder, 2004). Microorganisms degrade these compounds by using enzymes in their metabolism and can be useful in cleaning up contaminated sites (Alexander, 1999).

Microbial remediation of a hydrocarbon-contaminated site is accomplished with the help of a diverse group of microorganisms, particularly the indigenous bacteria present in soil (Osarumwense and Igiebor, 2018). These microorganisms can degrade a wide range of target constituents present in oily sludge (Barathi and Vasudevan, 2001; Mishra *et al.*, 2001). A large number of *Pseudomonas* strains capable of degrading PAHs have been isolated from soil and aquifers (Johnson *et al.*, 1996). Harder (2004) estimated that bioremediation accounts for 5 to 10 percent of all pollution treatment and has been used successfully in cleaning up the illegal dumping of used engine oil.

MATERIALS AND METHODS

Sample Collection

Soil samples were collected from an auto-mechanic workshop opposite the University of Benin, Benin City and were packed in sterile polybags and transferred to the laboratory for analysis.

Dilution of Sample

1g of soil sample was weighed and added into 9ml of distilled water. Then, 1ml from the sample was taking out and added into 9ml of distilled water. This step was continuously repeated until third dilution.

Isolation of Bacteria

Bacterial species were isolated from the collected soil samples by serial dilution and agar plating method wherein the soil sample was diluted from 10^{-1} to 10^{-3} dilutions, and the diluted soil samples were spread on sterile Nutrient agar plates. The inoculated plates were incubated at 37°C for 24 hours. Mixed cultures were obtained after incubation, labelled accordingly and purified by streaking on sterile nutrient agar plates. The purity of cultures was cross checked by gram staining procedures (Saroj and Keerti, 2013).

Staining and Biochemical Activities of Purified Cultures

In order to identify the purified cultures tentatively on the basis of Bergey's manual (Aneja, 2003), various staining and biochemical tests were performed namely Gram staining, Catalase test, Indole test, citrate utilization test, Urease test, Motility test, oxidase test, coagulase test, Glucose fermentation, fructose fermentation, and Lactose fermentation (Aneja, 2003).

Screening of Petroleum Degraders

The screening of petroleum degraders was done using the method of Osarumwense and Igiebor (2018).

RESULTS AND DISCUSSION

Table 1 shows the optical density (OD) at 600nm of two microorganisms (*P. aeruginosa* and *B. subtilis*) from day 0 to 14.

Table 1: Optical Density of *P. aeruginosa* and *B. subtilis* at 600nm.

Days	<i>P. aeruginosa</i>	<i>B. subtilis</i>
0	0	0
1	2.034	2.012
2	2.036	2.085
3	1.987	2.044
4	1.844	1.842
5	1.866	1.979
6	1.925	1.989
7	2.063	2.045
8	2.005	1.928
9	1.899	1.898
10	1.871	1.774
11	1.805	1.786
12	1.644	1.43
13	1.231	1.036
14	1.107	1.021

Table 2 above shows the total amount of hydrocarbon content (THC) degraded and their percentage degradation by the two

microorganisms from day 0 – 14. The total amount of hydrocarbon degraded on day 2 by *P. aeruginosa* was 0.088947 compared to the amount degraded by *B. subtilis* which was 0.04709, on the same day. The percentage degraded on day 6 by both organisms were significantly different, as percentage degraded by *P. aeruginosa* was 35.90582 and that degraded by *B. subtilis* was 29.82341. However, there were no significant differences between the amounts and percentages degraded by both organisms on day 10.

DISCUSSION

There are many reports on the degradation of hydrocarbon pollutants by different bacteria. Several bacteria are even known to feed exclusively on hydrocarbons (Yakimov *et al.*, 2007). Bacteria with the ability to degrade hydrocarbons are named hydrocarbon-degrading bacteria. Kafilzadeh *et al.* (2011) isolated 80 bacteria strains which belonged to 10 genus, which are; *Bacillus*, *Corynebacterium*, *Staphylococcus*, *Streptococcus*, *Shigella*, *Alcaligenes*, *Acinetobacter*, *Escherichia*, *Klebsiella* and *Enterobacter* and *Bacillus* were the best hydrocarbon degrading bacteria.

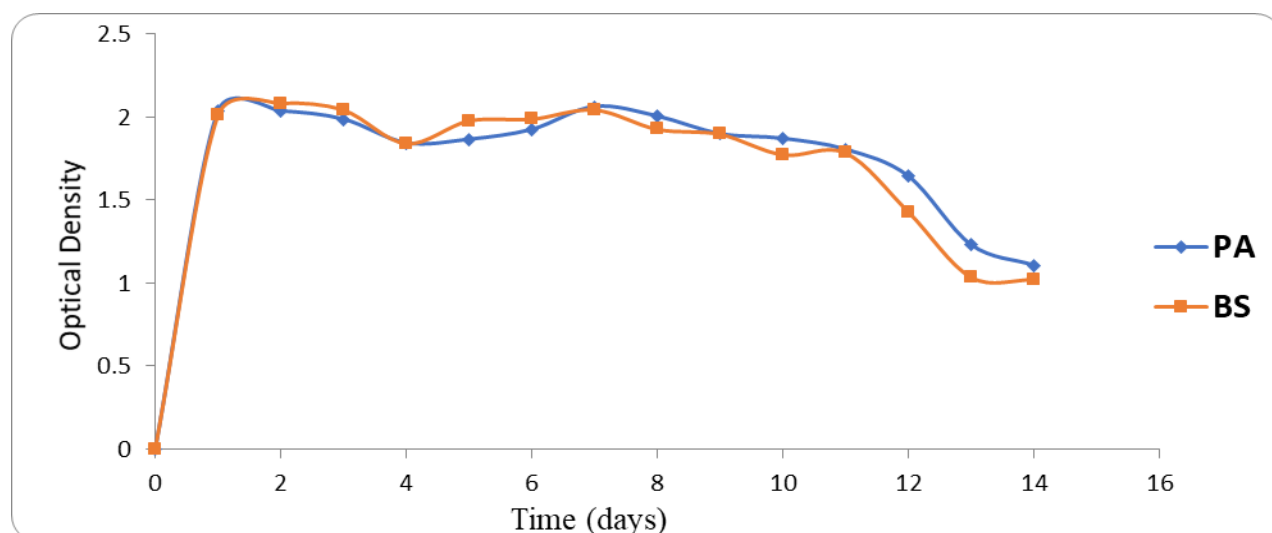


Figure 1: Variation of Optical Density with Time During the Degradation of Used Oil by *P. aeruginosa* (PA) and *B. subtilis* (BS).

Table 2: Total Amount of Hydrocarbon Content Degraded by *P. aeruginosa* and *B. subtilis*.

AMOUNT OF TOTAL HYDROCARBON CONTENT DEGRADED						
DAYS	<i>P. aeruginosa</i>			<i>B. subtilis</i>		
	THC (mg/kg)	Amount degraded (mg/kg)	% degraded	THC (mg/kg)	Amount degraded (mg/kg)	% degraded
0	152.90	0	0	152.90	0	0
2	139.30	0.088947	8.894702	145.70	0.04709	4.70896
4	125.60	0.178548	17.85481	118.10	0.2276	22.75997
6	98.00	0.359058	35.90582	107.30	0.298234	29.82341
8	86.00	0.437541	43.75409	77.60	0.492479	49.24787
10	72.50	0.525834	52.58339	73.20	0.521256	52.12557
12	71.80	0.530412	53.0412	58.80	0.615435	61.54349
14	71.30	0.533682	53.36821	57.30	0.625245	62.52453

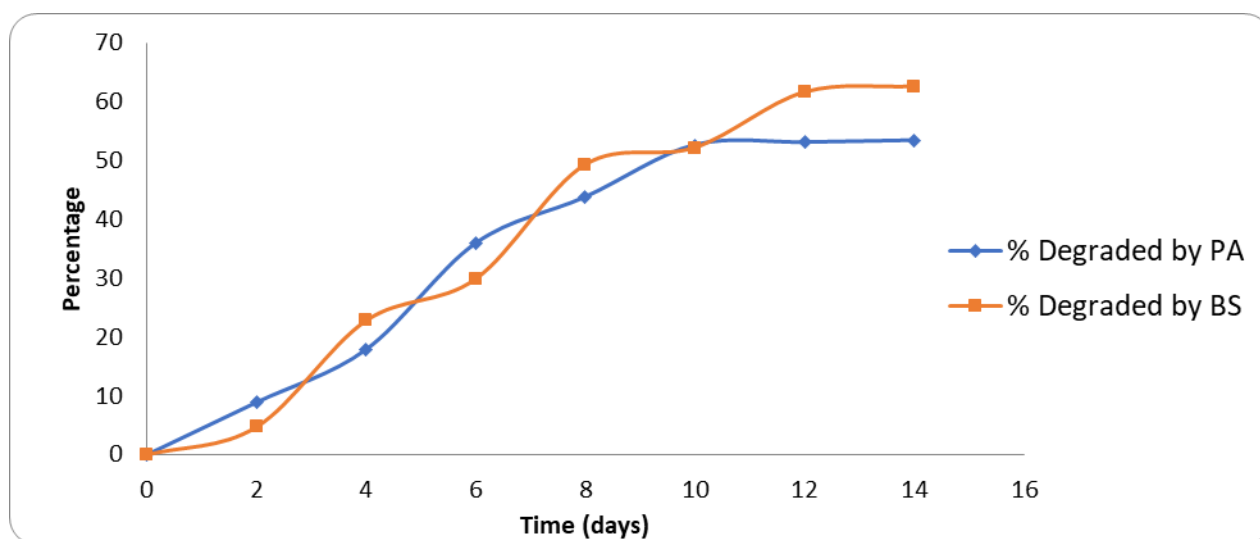


Figure 2: Percentage Degradation of Used Oil by *P. aeruginosa* (PA) and *B. subtilis* (BS).

Bacteria strains that are able to degrade aromatic hydrocarbons have been repeatedly isolated, mainly from soil. These are usually gram-negative bacteria most of them belong to the genus *Pseudomonas*. The biodegradative pathways have also been reported in bacteria from the genera *Mycobacterium*, *Corynebacterium*, *Aeromonas*, *Rhodococcus* and *Bacillus* (Mrozik *et al.*, 2003). Although many bacteria are able to metabolize organic pollutants, a single bacterium

does not possess the enzymatic capability to degrade all or even most of the organic compounds in a polluted soil.

Mixed microbial communities have the most powerful biodegradative potential because the genetic information of more than one organism is necessary to degrade the complex mixtures of organic compounds present in contaminated areas (Fritsche and Hofrichter, 2005). The

polluted soil samples were enriched with the hydrocarbon degrading bacteria and a total of 9 bacteria were isolated from the oil contaminated soil. These isolates were purified from the soil sample on the basis of colony, morphology, texture, growth. Biochemical tests like starch hydrolysis, urease production, citrate, coagulase and catalase test.

The indigenous bacteria isolated from hydrocarbon polluted soil were similar to the ones isolated by Osarumwense and Igebor (2018) which include; *Streptococcus* spp., *Escherichia coli*, *Micrococcus latus*, *Clostridium* spp., *Bacillus subtilis*, *Pseudomonas aeruginosa*, *Staphylococcus epidermidis*, *Proteus* spp. and *Staphylococcus aureus*. From the studies, bacteria identification using biochemical test revealed that *S. aureus* was mostly present in the soils polluted with hydrocarbons. The growth of the organism may be attributed to the fact that the enzymes are stable and metabolically active at 37°C.

Pseudomonas and *Bacillus* species are the most common bacteria hydrocarbon-degraders reported in the literatures (Barathi and Vasudevan, 2001; Bhattacharya *et al.*, 2002; Pokethitiyook *et al.*, 2003; Van Hamme *et al.*, 2003). This is also supported by the present studies that *P. aeruginosa* was still able to thrive well at 45°C. Changes in soil colour were observed after a period of time and may be as a result of the biodegradation potentials of the organisms.

Christopher and Christopher (2004) reported a sequential change of the composition of the oil degrading bacteria over a period of time in oil contaminated soil samples. Komukai-Nakamura *et al.* (1996) reported the sequential degradation of Arabian light crude oil by *Acinetobacter* sp T4 and *Pseudomonas putida* PB4. Successful removal of hydrocarbon by the addition of bacteria had been reported by Osarumwense and Igebor (2018). Microorganisms have developed the capabilities to protect themselves from toxicity by various mechanisms, such as adsorption, uptake, methylation, oxidation and reduction.

CONCLUSION

Microbial activities are very important for the renewal of our environment and maintenance of the global carbon cycle. These activities amidst

other substances can be degraded or transformed by microorganisms. However, in most cases these degradabilities which were estimated in the laboratory by using selected cultures and under ideal growth conditions, have shown that *P. aeruginosa* and *B. subtilis* have the potentials to degrade hydrocarbon polluted soil.

REFERENCES

1. Alexander, M. 1999. *Biodegradation and Bioremediation (2nd edition)*. Academic Press: San Diego, CA.
2. Aneja, K.R. 2003. *Experiments in Microbiology, Plant Pathology and Biotechnology*. New Age International (p). Ltd., Publishers: New Delhi, India.
3. Anoliefo, G.O. and B.O. Edegbai. 2000. "Effect of Spent Engine Oil as a Soil Contaminant on the growth of two Egg Plant Species, *Selenium belonging* L. and *S. inanes* L." *Journal of Agricultural, Forestry and Fisheries*; 1:21 – 25.
4. Barathi, S. and N. Vasudevan. 2001. "Utilization of Petroleum Hydrocarbons by *Pseudomonas fluorescens* isolated from a Petroleum–Contaminated Soil". *Environment International*. 26: 413 – 416.
5. Bhattacharya, D., P.M. Sarma, S. Krishnan, S. Mishra, and B. Lal. 2002. "Evaluation of Genetic Diversity among *Pseudomonas citronellolis* strains Isolated from Oily Sludge–Contaminated Sites". *Applied Environmental Microbiology*. 69(3):1435–1441.
6. Blodgett, W.C. 2001. "Water–Soluble Mutagen Production during the Bioremediation of Oil–Contaminated Soil". *Florida Scientist*. 60(1):28–36.
7. Boonchan, S., M.L. Britz. and G.A. Stanley. 2000. "Degradation and Mineralization of High–Molecular Weight Polycyclic Aromatic Hydrocarbons by Defined Fungal–Bacterial Cocultures". *Applied Environmental Microbiology*. 66(3): 1007–1019.
8. Butler, C.S. and J.R. Maso. 1997. "Structure–Function Analysis of the Bacterial Aromatic Ring–Hydroxylating Dioxygenases". *Advanced Microbial Physiology*. 38: 47–84.
9. Christopher, W.K. and L.K. Christopher. 2004. "Bacterial Succession in a Petroleum Land Treatment Unit". *Applied Environmental Microbiology*. 70(3):1777 – 1785.

10. Fritsche, W. and M. Hofrichter. 2005. "Aerobic Degradation of Recalcitrant Organic Compounds by Microorganisms". In: *Environmental Biotechnology: Concepts and Applications*. H.J. Jördening and J. Winter (eds.). Wiley-VCH Verlag GmbH & Co.: Berlin, Germany. doi: 10.1002/3527604 286.ch7.
11. Hagwell, I.S., L.M. Delfino, and J.J. Rao. 1992. "Partitioning of Polycyclic Aromatic Hydrocarbons from Oil into Water". *Environmental Science and Technology*. 26:2104– 2110.
12. Harder, E. 2004. *Bioremediation of Engine Oil*. Little Flower Academy: Dallas, TX.
13. Igiebor, F.A., J.O. Osarumwense, B.O. Obinyan, and P.C. Okoye. 2017. "Isolation and Identification of Indigenous Hydrocarbon Tolerant Fungi from Soil Contaminated with Biodiesel in Benin City, Nigeria". *International Journal of Agriculture and Environmental Science*. 4 (6): 47-50.
14. Ikhajiagbe, B. and G.O. Anoliefo. 2011. "Impact of Substrate Amendment on the Polyaromatic Hydrocarbon Content of a Five-Month Old Waste Engine Oil Polluted Soil". *African Journal of Environmental Sciences and Technology*. 5(10):769 – 777.
15. Johnson, K., S. Anderson, and C.S. Jacobson. 1996. "Phenotypic and Genotypic Characterization of Phenanthrene-Degrading Fluorescent *Pseudomonas biovars*". *Applied Environmental Microbiology*. 62: 3818–3825.
16. Kafilzadeh, F., P. Sahragard, H. Jamali and Y. Tahery. 2011. "Isolation and Identification of Hydrocarbons Degrading Bacteria in Soil Around Shiraz Refinery". *African Journal of Microbiology Research*. 4(19): 3084 – 3089.
17. Komukai-Nakamura, S., K. Sugiura, Y. Yamauchi-inomata, H. Toki, K. Venkateswaran, S. Yamamoto, H. Tanaka, and S. Harayama. 1996. "Construction of Bacterial Consortia that Degrade Arabian Light Crude Oil". *Journal of Fermentation and Bioengineering*. 82: 570 – 574.
18. Mishra, S.J., R.C. Jyot, and B. Kuhad. 2001. "Evaluation of Inoculums Addition to Stimulate in situ Bioremediation of Oily-Sludge-Contaminated Soil". *Applied Environmental Microbiology*. 67(4): 1675–1681.
19. Mroziak, A., Z. Piotrowska-Seget, and S. Labuzek. 2003. "Bacterial Degradation and Bioremediation of Polycyclic Aromatic Hydrocarbons". *Polish Journal of Environmental Studies*. 12(1): 15 – 25.
20. Odu, C.T.I. 1981. "Degradation and Weathering of Crude Oil under Tropical Condition". In: *Proceeding of an International Seminar on the Petroleum Industry and the Nigeria Environment*. Petroleum Training Institute: Warri, Nigeria.
21. Osarumwense, J.O. and F.A. Igiebor. 2018. "Assessment of Indigenous Bacteria from Biodiesel Effluents Contaminated Site". *Journal of Applied Sciences and Environmental Management*. 22(2): 157-160.
22. Pokethitiyook, P., A. Sungpetch, S. Upathame, and M. Kruatrachue. 2003. "Enhancement of *Acinetobacter calcoaceticus* in Biodegradation of Tapis Crude Oil". *Applied Environmental Microbiology*. 42: 1–10.
23. Saroj, A. and D. Keerti. 2013. "Isolation and Characterization of Hydrocarbon Degrading Microorganisms from Petroleum Oil Contaminated Soil Sites". *Bulletin of Environmental and Scientific Research*. 2(4): 5 – 10
24. Udo, E.J. and A.A. Fayemi. 1975. "The Effect of Oil Pollution on Germination, Growth and Nutrient Uptake of Corn". *Journal of Environmental Quality*. 4:537 – 540.
25. Van Hamme, J.D., A. Singh, and O.P. Ward. 2003. "Recent Advances in Petroleum Microbiology. Microbiology Molecular Biology Review". 67(4): 503–549.
26. Yakimov, M.M., K.N. Timmis, and P.N. Golyshin. 2007. "Obligate Oil-Degrading Marine Bacteria". *Current Opinion in Biotechnology*. 18(3) 257 – 266.

ABOUT THE AUTHORS

Dr. Jude O. Osarumwense, is a Lecturer in the Department of Science Laboratory Technology, Faculty of Life Sciences, University of Benin, Benin City, Nigeria. He holds a Ph.D. degree in Chemical Engineering from the University of Benin. His research interests are in renewable energy utilization and environmental engineering. He is a Registered Engineer, a member of the Nigerian Society of Chemical Engineers, and an Associate Member of the Nigerian Institute of Science Laboratory Technology (ANISLT).

Francis A. Igiebor, is a Researcher at Wellspring University and University of Benin, Nigeria. He is a registered Laboratory Technologist, an Associate member of the Nigerian Institute of Science Laboratory Technology (ANISLT), and a Member of Microbiology Society of Nigeria (MSN). He holds a Master of Science (M.Sc.) degree in Environmental Plant Physiology from the

University of Benin. His research interests are in remediation and environmental sustainability.

Daniel E. Idahosa, is a research student supervised by Dr. Osarumwense in the Department of Science Laboratory Technology, University of Benin, Nigeria. He holds a B.Sc. degree in Science Laboratory Technology (Microbiology Techniques) and currently is an Associate Member of the Nigerian Institute of Science Laboratory Technology (ANISLT).

SUGGESTED CITATION

Osarumwense, J.O., F.A. Igiebor, and D.E. Idahosa. 2019. "Isolation, Characterization and Identification of Bacterial isolates from Auto-mechanic Workshop contaminated with Hydrocarbon". *Pacific Journal of Science and Technology*. 20(1):349-355.

